

---

ANNALES  
UNIVERSITATIS MARIAE CURIE-SKŁODOWSKA  
LUBLIN – POLONIA

VOL. LXII (2)

SECTIO DD

2007

---

Zakład Chorób Ryb i Biologii Akademii Rolniczej w Lublinie  
ul. Akademicka 12, 20-033 Lublin  
e-mail: leszek.guz@ar.lublin.pl

\*Katedra Technologii Przemysłu Rolno-Spożywczego i Przechowalnictwa  
ul. Skromna 8, 20-704 Lublin  
e-mail: awasko@tlen.pl

LESZEK GUZ, ADAM WAŚKO\*, ANTONINA Sopińska,  
MARIAN JAMROZ

### **Extracellular protease production by *Moritella viscosa***

Wytwarzanie pozakomórkowych proteaz przez *Moritella viscosa*

**Summary.** *Moritella viscosa* (*Vibrio viscosus*) is the causative agent of winter ulcers in farmed salmonids, and bacteria have been isolated from various marine fish species. The protease activity at 6°C and 16°C of *M. viscosa* strain were studied. Substrate gel electrophoresis with gelatin and casein as the substrates showed that *M. viscosa* strain had two different proteinase bands, 73.5 and 67.5 kDa.

**Key words:** *Moritella viscosa*, winter ulcer, fish

#### INTRODUCTION

*Moritella viscosa* (*Vibrio viscosus*) is a psychrotrophic bacterium that has been found in winter ulcers in salmonids in Norway, Iceland, Scotland, Denmark, Ireland, and Canada [Lunder 1992, Lunder *et al.* 1995, 2000, Benediksdottir *et al.* 1998, Bruno *et al.* 1998, Lillehaug *et al.* 2003, Håstein *et al.* 2005], first described as *Vibrio viscosus* [Lunder *et al.* 2000]. Affected farmed fish, described by Lunder [1992], Benediksdottir *et al.* [1998] and Bruno *et al.* [1993] were lethargic, dark in colour, showed gill pallor, and exophthalmos. Extracellular, epidermal ulcers were recorded on the scale-covered areas and ranged in diameter from 0.5 to 2.0 cm. The ulcers were superficial or penetrated to the musculature. The disease occurs most frequently at low temperatures, thus explaining the name 'winter ulcer' ('vintersår' in Norwegian). In Norway the disease occurs mainly in February, March and April, and the fish usually recover when the water temperature rises above 8°C. *Moritella viscosa* infections have been detected in various fish species in the field, including salmon, cod, plaice, rainbow trout, and lumpfish [Benediksdottir *et al.* 2000, Lunder *et al.* 2000, Colquhoun *et al.* 2004, Gudmundsdóttir *et al.* 2006].

The aim of this study was to investigate the effect of incubation temperature upon the composition of extracellular proteases produced by *M. viscosa* strain.

## MATERIAL AND METHODS

**Bacteria and growth conditions.** *Moritella viscosa* strain was isolated from Atlantic salmon (*Salmo salar*), and was kindly provided from the collection at the Institute for Experimental Pathology, University of Iceland, Keldur, Reykjavik. The bacteria were cultured in tryptic soy agar containing 2% NaCl (TSA-2). The agar plates were incubated 48 hour at 16°C. For production of extracellular proteases bacteria were grown in *V. viscosus* medium (VM) for 24 h at 6°C and 18°C under constant shaking (180 RPM). *Vibrio viscosus* medium: MgSO<sub>4</sub> · 7 H<sub>2</sub>O – 1 g/l, yeast extract – 20 g/l, NaCl – 12 g/l, KH<sub>2</sub>PO<sub>4</sub> – 2 g/l, pH – 7,3.

**Antibiotic susceptibility tests.** The antibiotic susceptibility test was determined according to the procedure described by Ayres *et al.* [1999], with slight modifications. Briefly, antibiotic discs were purchased from Oxoid and consisted of amoxicilin 10 µg, oxolinic acid 2 µg, oxytetracycline 30 µg. Diagnostic sensitivity test agar (TSA-2) plates poured, and allowed to set. The cell suspension (0.25 ml) was pipeted onto the surface of each plate and spread evenly with a dry sterile swab. The inoculated plates were allowed to dry for 2 h, antibiotic discs placed onto the agar surface and the plates were incubated at 16°C for 24 h and the zones of inhibition measured.

**Biochemical tests.** Biochemical testing was carried out using API Zym test strips (bioMérieux) at 18°C and recorded after 72 h incubation.

**Measurement of proteolytic activity of ECPs.** Protein levels of ECPs solutions were determined using the Sigma protein assay kit with bovine albumin as standard.

**Caseinase activity.** The caseinase activity was determined by the azocasein procedure described by Leung and Stevenson [1988], with slight Mateos *et al.* [1993] and the authors' own modifications. Briefly, the reaction mixture consisted of 0.1 ml of a 10% (w/v) Azocasein solution (Sigma), 0.1 ml of supernatant fluid sample and 2.3 ml of 0.1 mol l<sup>-1</sup> sodium phosphate buffer, pH 7.2 and was incubated at 20°C for 24 h. The reaction was stopped with 2.5 ml of 10% (w/v) trichloroacetic acid (TCA), and after 30 min at room temperature, the precipitation was removed by centrifugation. Equal volumes of supernatant fluid and NaOH 1 mol l<sup>-1</sup> were mixed and absorbance was read at 450 nm. TCA was added to the blank before incubation.

**Elastase activity.** The caseinase activity was determined by the azocasein procedure described by Bjorn *et al.* [1979], with slight modifications by Mateos *et al.* [1993] and our own. Briefly, one ml of culture supernatant fluids was added to 2 ml of Tris-maleate buffer (0.1 mol l<sup>-1</sup>, pH 7.0) supplemented with CaCl<sub>2</sub> (0.001 mol l<sup>-1</sup>) containing 10 mg of elactin-Congo red. The mixture was incubated at 20°C for 24 h and the reaction was stopped by the addition of 2 ml of sodium phosphate buffer (0.7 mol l<sup>-1</sup>, pH 6.0). The precipitate was removed by centrifugation. The blank consisted of 3 ml of the buffer containing 10 mg of elastin-Congo red. Elastase activity was determined by reading absorbance of the supernatant fluid at 495 nm.

**SDS-PAGE.** Proteolytic activity was visualized in 12% SDS-polyacrylamide gels [Laemmli 1970] by incorporating 0.1% sodium caseinate or gelatin, according to the method described by Mateos *et al.* [1993]. The SDS was removed by shaking the gels in 2.5% Triton X-100 (Sigma) for 1 h and then the gels were incubated in glycine 0.1 mol l<sup>-1</sup> pH 8.3 overnight at 28°C to allow proteolysis to take place. Protease activity was visible as clear bands on a blue background. The relative migration distance was compared with a standard set of low molecular-weight markers (BioRad), to yield apparent molecular-weight values.

## RESULTS AND DISCUSSION

*Moritella viscosa* (previously *Vibrio viscosus*) has received increased attention as a cause of winter ulcer [Bruno *et al.* 1998, Benediktsdóttir *et al.* 2000, Lunder *et al.* 2000, Gudmundsdóttir *et al.* 2006] in farmed salmonids, cod and various other marine fish species, in several countries around the North Atlantic [Colquhoun *et al.* 2004, Björnsdóttir *et al.* 2004]. Winter ulcer is a disease affecting fish in sea water, most frequently during the winter months, when the temperature is below 10°C [Larsen and Pedersen 1999]. In Norway the disease occurs mainly in February, March and April and the fish usually recover when the water temperature rises above 8°C. Diseased fish show characteristic skin ulcers of varying size and depth. The mortality is usually low, from 0 to 10% during the outbreak, but the disease has economic significance due to lowered quality of the fish [Lunder 1992].

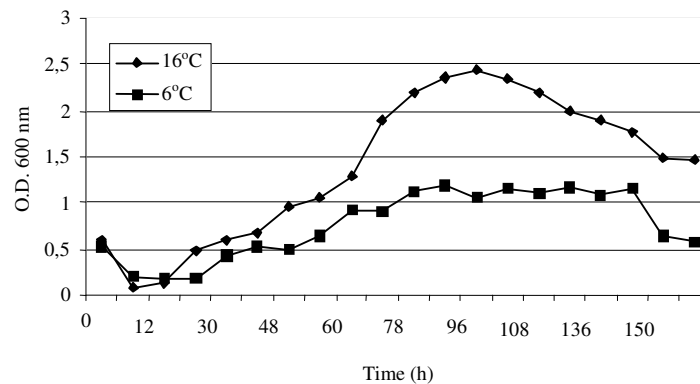


Fig. 1. Effect of temperature on growth of *Moritella viscosa* strain Rys. 1. Wpływ temperatury na wzrost *Moritella viscosa*

The *M. viscosa* cultures were sensitive to amoxicillin, oxolinic acid and oxytetracycline. The biochemical properties of the bacterium corresponded with those described for a novel *Vibrio* sp. by Lunder [1992] named *Vibrio viscosus*, and Benediktsdóttir *et al.* [2000]. Strains of *M. viscosa* are Gram-negative, short non-sporeforming rods that are motile by means of a single polar flagellum. The bacteria are oxidase and catalase positive, produced acid from glucose. They do not produce acid from D-arabinose, L-arabinose, cellobiose, glycerol, inositol, lactose, mannitol, melibiose, rhamnose, salicin, sorbitol, sucrose, and trehalose. *M. viscosa* strains are negative in arginine dihydrolase, ornithine decarboxylase and usually positive in lysine decarboxylase. Casein, gelatin, DNA, Tween 20, Tween 80 and starch are hydrolyzed, but neither alginate nor esculin are hydrolyzed. The Voges-Proscauer test is negative. The bacterium is susceptible to 2,4-diamino-6,7-diisopropylpteridine (vibriostatic agent, 0/129) [Lunder 1992, Bruno *et al.* 1998, Lunder *et al.* 2000, Benediktsdóttir *et al.* 2000]. Moreover, the production of lysine decarboxylase and the production of acid from maltose and mannose are different, depending on subgroups [Benediktsdóttir *et al.* 2000].

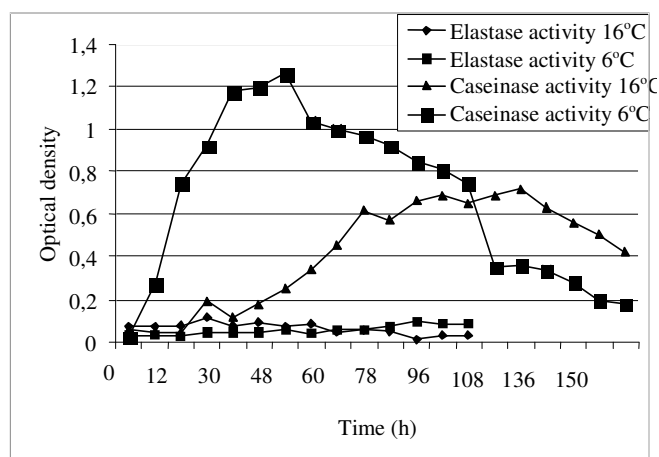


Fig. 2. Exoprotease production by *Moritella viscosa*  
Rys. 2. Wytwarzanie proteaz przez *Moritella viscosa*

Table 1. Biochemical characteristics of *Moritella viscosa* using API Zym kit from bioMérieux.  
Results recorded at 18°C after 72 h incubation  
Tabela 1. Biochemiczna charakterystyka *Moritella viscosa* testem API Zym z firmy bioMérieux.  
Wyniki pochodzą z 18°C po 72 godz. inkubacji

Enzyme	Activity mark of <i>Moritella viscosa</i> (at 18°C/6°C)*		
	Log phase	Stationary phase	Death phase
Phosphatase alkaline	0/3	0/5	0/5
Esterase (C 4)	1/1	2/1	2/1
Esterase Lipase (C 8)	3/3	4/3	4/3
Lipase (C 14)	2/0	2/1	1/1
Leucine arylamidase	4/1	5/1	4/3
Valine arylamidase	2/0	3/0	2/0
Cystine arylamidase	0/0	1/0	0/1
Trypsin	0/0	0/0	0/0
Chymotrypsin	1/1	2/1	2/1
Phosphatase acid	1/3	1/3	1/5
Naphthol-AS-BI-phosphohydrolase	1/1	1/1	1/1
α-galactosidase	0/0	0/0	0/0
β-galactosidase	0/0	0/0	0/2
β-glucuronidase	0/0	0/0	0/0
α-glucosidase	0/1	0/1	0/1
β-glucosidase	3/3	5/3	5/5
N-acetyl-β glucosaminidase	3/1	5/1	5/1
α-mannosidase	0/0	0/0	0/0
α-fucosidase	0/0	0/0	0/0

\*Quantity of hydrolysed substrate: 0 – 0 nM (nanomole); 1 – 5 nM; 2 – 10 nM; 3 – 20 nM; 4 – 30 nM; 5 - > 40 nM

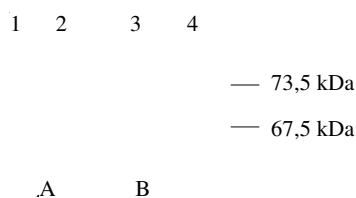


Fig. 3. Substrate SDS-PAGE analysis comparing extracellular products (ECPs) of *Moritella viscosa* measured in the middle of log phase: A – casein SDS-PAGE; B – gelatin SDS-PAGE; 1 and 3–6°C; 2 and 4–16°C

Rys. 3. Porównawcza analiza produktów zewnątrzkomórkowych (ECPs) *Moritella viscosa* mierzona w połowie fazy logarytmicznego wzrostu: A – kazeina SDS-PAGE; B – żelatyna SDS-PAGE; 1 i 3–6°C; 2 i 4–16°C

The observation that winter ulcer occurs mainly when water temperatures are low reflects the decreased reparative and regenerative ability that fish possesses at these temperatures [Roberts 1975]. It is well known that temperature is an important factor controlling the rate of development of microbial populations, a modulation of enzyme synthesis by the growth temperature having been observed in several microorganisms [Herdendeen *et al.* 1979, Gügi *et al.* 1991, Mateos *et al.* 1993]. In our study, short, curved, Gram-negative staining rods were recorded on TSA-2 medium. The colonies were translucent, creamy yellow in appearance and viscous when touched with a loop. The bacteria were grown at 6°C and 16°C for 162 h. Figure 1 shows that the growth of bacteria was considerably slower at 6°C than at 16°C. The effect of incubation time and temperature on protease activity of *M. viscosa* ECPs is shown in Fig. 2. At 16°C the bacterial growth reached its maximum effect after 96 h of incubation time. The lowest caseinase activity was observed in the supernatant fluid of culture grown at 16°C for 124 h. Cultures grown at 6°C showed that lowest bacterial growth, but highest caseinase activity reached its maximum after 48 h of incubation time were observed. Moreover, the elastase activities were not observed at either 6°C or 16°C. Biochemical properties of strain cultured at 6°C and 16°C (at log phase, stationary phase and death phase) are reported in Table 1.

Substrate gel electrophoresis with gelatin or casein as the substrates showed that *M. viscosa* strain produced two different bands showing proteolytic activity. Neither of them were affected by the treatment with  $\beta$ -mercaptoethanol. Substrate SDS-PAGE patterns of ECPs representative for all caseinase and gelatinase profiles are shown in Fig. 3. Substrate gel electrophoresis with gelatin and casein as the substrates showed that *M. viscosa* strain had two different proteinase bands, 73.5 kDa and 67.5 kDa. The influence of temperature of incubation on the types of produced proteases was scarce so all the proteases detected were produced at both 16°C and 6°C.

## REFERENCES

- Ayres H.M., Furr J.R., Russel A.D. 1999: Effect of permeabilizers on antibiotic sensitivity of *Pseudomonas aeruginosa*. Lett. Appl. Microbiol. 28, 13–16.
- Benediktsdóttir E., Helgason S., Sigurjónsdóttir H. 1998: *Vibrio* spp. isolated from salmonids with shallow skin lesions and reared at low temperature. J. Fish Dis. 21, 19–28.
- Benediktsdóttir E., Verdonck L., Spröer C., Helgason S., Swings J. 2000: Characterization of *Vibrio viscosus* and *Vibrio wodanis* isolated at different geographical locations: a proposal for reclassification of *Vibrio viscosus* as *Moritella viscosa* comb. nov. Int. J. Syst. Evolution. Microbiol. 50, 479–488.
- Bjorn M.J., Sokol P.A., Iglewski B.H. 1979: Influence of iron on yields of extracellular products in *Pseudomonas aeruginosa* cultures. J. Bacteriol. 138, 193–200.
- Björnsdóttir B., Gudmundsdóttir S., Bambir S.H., Magnadóttir B., Gudmundsdóttir B.K. 2004: Experimental infection of turbot, *Scophthalmus maximus* (L.), by *Moritella viscosa*, vaccination effort and vaccine-induced side-effects. J. Fish Dis. 27, 645–655.
- Bruno D.W., Griffiths J., Petrie J., Hastings T.S. 1998: *Vibrio viscosus* in farmed Atlantic salmon, *Salmo salar* in Scotland: field and experimental observations. Dis. Aq. Org. 34, 161–166.
- Colquhoun D.J., Hovland H., Helleberg H., Haug T., Nilsen H. 2004: *Moritella viscosa* isolated from farmed Atlantic cod (*Gadus morhua*). Bull. Eur. Ass. Fish Pathol. 24, 109–114.
- Gudmundsdóttir B.K., Björnsdóttir B., Gudmundsdóttir S., Bambir S.H. 2006: A comparative study of susceptibility and induced pathology of cod, *Gadus morhua* (L.), and halibut, *Hippoglossus hippoglossus* (L.), following experimental infection with *Moritella viscosa*. J. Fish Dis. 29, 481–487.
- Gügi B., Orange N., Hellio F., Burini J.F., Guillen C., Leriche F., Guespin-Michel J.F. 1991: Effect of growth temperature on several exported enzyme activities in the psychrotrophic bacterium *Pseudomonas fluorescens*. J. Bacteriol. 173, 3814–3820.
- Herendeen S.L., Van Bogelen R.A., Neidhardt F.C. 1979: Levels of major proteins of *Escherichia coli* during growth at different temperatures. J. Bacteriol. 139, 185–194.
- Håstein T., Gudding R., Evensen Ø. 2005: Bacterial vaccines for fish – an update of the current situation worldwide. Dev. Biol. 121, 55–74.
- Laemmli U.K. 1970: Cleavage of structural proteins during assembly of the head of bacteriophage T4. Nature (London) 227, 680–685.
- Leung K.Y., Stevenson R.M.W. 1988: Tn 5 induced protease-deficient strains of *Aeromonas hydrophila* with reduced virulence for fish. Inf. Immun. 56, 2639–2644.
- Lillehaug A., Lunestad B.T., Grave K. 2003: Epidemiology of bacterial diseases in Norwegian aquaculture – a description based on antibiotic prescription data for the ten-year period 1991 to 2000. Dis. Aq. Org. 53, 115–125.
- Lunder T.: Winter ulcer in Atlantic salmon. PhD Thesis, Norwegian College of Veterinary Medicine, Oslo 1992.
- Lunder T., Evensen Ø., Holstad G., Håstein T. 1995: ‘Winter ulcer’ in the Atlantic salmon *Salmo salar*. Pathological and biochemical investigations and transmission experiments. Dis. aquat. Org. 23, 39–49.
- Lunder T., Sørum H., Holstad G., Steigerwalt A.G., Mowinckel P., Brenner D.J. 2000: Phenotypic and genotypic characterization of *Vibrio viscosus* sp. nov. and *Vibrio wodanis* sp. nov. isolated from Atlantic salmon (*Salmo salar*) with ‘winter ulcer’. Int. J. Syst. Evol. Microbiol. 50, 427–450.

- Mateos D., Anguita J., Naharro G., Paniagua C. 1993: Influence of growth temperature on the production of extracellular virulence factors and pathogenicity of environmental and human strains of *Aeromonas hydrophila*. J. Appl. Bacteriol. 74, 111–118.
- Santos J.A., González C.J., López-Díaz T.M., García-López M.L., Otero A. 1996. Extracellular protease production by dairy strains of *Aeromonas hydrophila* as affected by growth media and incubation temperature. Food Microbiol. 13, 47–51.

**Streszczenie.** ‘Winter ulcer’ jest chorobą głównie ryb łososiowatych, wywoływaną przez bakterie *Moritella viscosa* (dawniej *Vibrio viscosus*). Chorobę notowano w Norwegii, Islandii, Szkocji, Danii, Irlandii i Kanadzie. Występuje w zimnych porach roku, gdy temperatura wody spada poniżej 10°C. Śmiertelność ryb jest niewielka, ale ze względu na niższą jakość mięsa przynosi znaczne straty ekonomiczne. W niniejszej pracy zbadano wzrost bakterii *M. viscosa* w temperaturze 16°C – optymalnej do wzrostu oraz w temperaturze 6°C. Zbadano również aktywność kazeinazową i elastazową oraz wykonano analizę zymograficzną proteaz (SDS-PAGE), wykorzystując kazeinę i żelatynę jako substraty. Wykazano, że badany szczep wytwarza dwie proteazy o aktywności żelatynazowej i dwie proteazy o aktywności kazeinazowej.

**Słowa kluczowe:** *Moritella viscosa*, ‘winter ulcer’, ryby